Synthesis of 2'-C-cyano-2'-deoxy- And 2'-C-cyano-2',3'-dideoxy-β-Darabinofuranosyl Nucleosides

Sonsoles Velázquez and María-José Camarasa*

Instituto de Química Médica, Juan de la Cierva, 3. 28006 Madrid, Spain

(Received in UK 13 January 1992)

Abstract.—A series of 2'-C-cyano-2'-deoxy- and 2'-C-cyano-2', 3'-dideoxy arabinofuranosyl nucleosides have been prepared by reaction of the corresponding 2'-ulosyl- and 3'-deoxy-2'-ulosyl nucleosides with sodium cyanide followed by 2'-deoxygenation of the cyanohydrins formed and removal of the protecting groups of the sugar moiety.

INTRODUCTION

2',3'-Dideoxynucleosides, including AZT and ddI are potent and selective anti-HIV agents, which seem to act through inhibition of Reverse Transcriptase.^{1,2,3} However, despite its benefits,⁴ severe side effects with AZT treatment are described, mainly associated with bone marrow toxicity.^{5,6} In an attempt to increase the antiviral activity and to decrease cytotoxicity of AZT, several analogs of AZT and 2'3'-dideoxynucleosides have been synthesized.^{7,8} One approach in this field consisted of the synthesis of 2',3'-ddN derivatives in which the 3'-OH has been replaced by a 3'-substituent, which mimicks the stereoelectronic properties of the 3'-OH or the azido group of AZT.⁷ 3'-C-Branched-chain sugar nucleosides bearing at the branching point groups that mimick the azido or the OH groups [CN, ethynyl, allyl, propargyl, cyanomethyl and other three atom fragments (C-C=X, C-C=X)] were considered good candidates to inhibit the Reverse Transcriptase of HIV.⁷

As a part of our program on the synthesis of branched-chain sugar nucleosides as inhibitors of HIV replication, we reported on the synthesis and the anti HIV-1 activity 9 of 3'-C-cyano branched chain nucleosides. Our approach 10,11 involved the reaction of pentofuranos-3'-ulosyl nucleosides with sodium cyanide followed by deoxygenation of the cyanohydrin thus formed. We have used this procedure for the synthesis of a variety of 3'-C-cyano-3'-deoxynucleoside analogues. 9,11 In the present paper we extend this procedure to the pentofuranos-2'-ulosyl nucleosides and prepare 2'-C-cyano-2'-deoxy- and 2'-C-cyano-2',3'-dideoxypentofuranosyl derivatives of uracil, cytosine and adenine.

RESULTS AND DISCUSSION

Oxidation of 4-N-monomethoxytrityl-1[3',5'-O-(tetraisopropyldisiloxan-1,3-diyl)- β -D-ribofuranosyl]cytosine¹² with CrO₃/pyridine/Ac₂O¹³ gave 4-N-monomethoxytrityl-1[3',5'-O-(tetraisopropyldisiloxan-1,3-diyl)- β -Derythro-pentofuranos-2'-ulosyl] cytosine (**1b**) in 72% yield. Treatment of the 2'-ketonucleosides of uracil **1a**¹³, 4-N-monomethoxytrityl (MMTr) cytosine 1b, and adenine $1c^{13}$ with sodium cyanide in a two-phase ethyl ether / water system, in the presence of sodium bicarbonate, gave in each case a mixture of the two epimeric 2'- cyanohydrins 2 and 3. These cyanohydrins, on standing in solution, reversed to the corresponding ketonucleosides 1, used as starting materials. Thus, they were not isolated, and were used as such, without further purification for the next step. Deoxygenation^{14,15} at the 2'-position of the mixtures of cyanohydrins 2 and 3 by treatment with (phenyloxy) thiocarbonyl chloride in the presence of 4-(dimethylamino)pyridine followed by reaction with tributyltin hydride in the presence of AIBN, afforded, stereoselectively, the 2'-C-cyano-2'-deoxy-arabinofuranosyl derivatives of uracil 4a, 4-N-MMTr cytosine 4b and adenine 4c, in 40%, 42% and 45% yield, respectively.



Deprotection of **4a-c** with tetrabutylammonium fluoride afforded the corresponding 2'-C-cyano- β -Darabino deprotected nucleosides of uracil **5a** in low yield (30%) and adenine **5c** (60% yield), unfortunately the corresponding deprotected derivative of cytosine (**5b**) was not obtained and only cytosine was isolated from the reaction mixture. The low yield obtained of the uracil derivative and the no isolation of the corresponding cytosine derivative, would be explained by the higher acidity of the H-2' proton, with respect to other 2'deoxynucleosides, due to the presence, in the same carbon atom, of the cyano group, which would lead to β elimination of the nucleobase.

This made us to try a different strategy for the synthesis of the 2'-C-cyano-2',3'-ddN which consisted of deoxygenation of the 3' position of the conveniently 2',5'-protected nucleosides and then introduction of the cyano group at 2'-position followed by 2'-deoxygenation, and removal of the 5'-protecting group of the nucleoside under acidic conditions in order to avoid the β -elimination of the nucleobase.

The 2',3'-dideoxy-2'-C-cyano analogues were prepared from the corresponding 3'-deoxy-2'ketonucleosides. Thus, the 2',5'-bis-O-silylated derivatives of uracil **6a**¹⁶, 4-N-MMTr-cytosine **6b** and 5'-MMTr-2'-O-silylated derivative of adenine **6d**¹⁶ were deoxygenated¹⁴ at 3'-position by treatment with N,Nthiocarbonyldiimidazol followed by reaction with tributyltinhydride in the presence of α, α' -AIBN to give the 3'deoxyderivatives **7a**, **7b** and **7d** in 73%, 52% and 75% yield, respectively.

Treatment of **7a-d** with tetrabutylammonium fluoride afforded the corresponding 2',5'-O-deprotected-3'deoxyuridine **8a**¹⁷, cytidine **8b** and the 5'-MMTr-2'-deprotected derivative of adenine **8d**.¹⁸ Reaction of **8a** and



8b with 1 eq. of t-butyldimethylsilyl chloride in dry pyridine gave the 5'-O-silylated nucleosides **9a** and **9b** in 72% and 81% yield, respectively. Oxidation of the 5'-O-protected nucleosides **9a**, **9b** and **8d** with $CrO_3/pyridine/Ac_2O^{13}$ afforded the 2'-ketonucleosides **10a**, **10b** and **10d** in 65%, 89% and 53% yield, respectively. **10a**, **10b** And **10d** were treated with sodium cyanide in ethyl ether/water, in the presence of sodium bicarbonate, to give a mixture of the epimeric nucleoside 2'-cyanohydrins **11** and **12**, further treatment of the mixture (**11+12**) with (phenyloxy)thiocarbonyl chloride and 4-(dimethylamino)pyridine followed by reaction with Bu₃SnH and AIBN, gave the 2'-C-cyano-2',3'-dideoxyderivatives **13a**, **13b** and **13d** in 50%, 36% and 25% yield, respectively.

Finally, deprotection of 13a and 13b with methanolic 0.1 M HCl gave the deprotected nucleosides of uracil 14a and cytosine 14c in 86% and 80% yield, respectively. Similarly, deprotection of the 5'-O-MMTr, group of 13d with 80% aqueous acetic acid afforded 14d in 70% yield.

The presence of the 2'-C-cyano-2'-deoxy grouping in nucleosides 4 and 5, and the 2'-C-cyano-2',3'dideoxy grouping in nucleosides 13 and 14 was established from their analytical and spectroscopic data. The IR spectra of 4a-c, 5a-c, 13a-d and 14a-d showed a band at 2220-2230 cm⁻¹ characteristic of CN group. The ¹HNMR spectra of 4a-c and 5a-c showed the presence of H-2' at δ 3.60-4.07, as a doublet of doublets for 4a, 4c and 5c or as a triplet for 4b and 5a, by coupling with H-1' and H-3'. The ¹³CNMR of 4a-c and 5a-c showed a signal at δ 115.29-117.14 assigned to the CN carbon atom, and a signal at δ 42.15-45.12 corresponding to the C-2'. On the other hand, the ¹HNMR spectra of 13a-d and 14a-d showed the presence of a multiplet corresponding to H-2' at δ 3.59-4.30 and two multiplets, overlapped in some cases, corresponding to the 2H-3' at δ 2.1-2.30. The ¹³CNMR of 13a-d and 14a-d showed the signal corresponding to the C-3' at δ 29.25-31.39.



The stereochemistry at C-2' of the arabino 2'-C-cyanonucleosides 4, 5, 13 and 14 was unequivocally determined by n.O.e. experiments, ^{19,20} the signal of H-1' was irradiated and the magnitude of n.O.e. at the protons of the sugar were observed (Fig 1). Thus, in compounds 4a-c and 5a-c irradiation of H-1' induced n.O.e. to H-2' [4a (21%), 4b (19%), 4c (14%), 5a (20%), 5c (19%)] and to H-4' [4a (1%), 4b (3%), 4c (4%), 5a (1%), 5c (2%)]. Similarly, irradiation of H-1' in 13a-d and 14a-d induced n.O.e. to H-2' [13a (10%), 13b (15%), 13d (10%), 14a (17%), 14c (16%), 14d (15%)] and to H-4' [13a (3%), 13b (4%), 13d (2%), 14a (4%), 14c (1%), 14d (1%)]. This demonstrates that protons H-1', H-2' and H-4' are all in the α -

face of the furanose ring and that no epimerization at C-2' had occurr during removal of the protecting groups of the final products $4 \rightarrow 5$ and $13 \rightarrow 14$.

The arabino configuration of compounds 4, 5, 13 and 14 resulted from 2'-deoxygenation of cyanohydrins (2 and 3) and (11 and 12), and can be explained by steric hindrance of the β -face of the furanose ring, which facilitates the approach of the tributyltin hydride from the less hindered α face of the molecule,¹⁴ opposite to the heterocyclic base at the anomeric position. The stereochemistry of the above reactions is in agreement with those observed in most 2'-radical deoxygenations.^{21,22,23}

The compounds described above have been tested for antiviral activity against HIV-1, but none of them showed any significant activity, nor toxicity. Further tests of these compounds in other viruses are in progress.

EXPERIMENTAL

Microanalyses were obtained with a Heareus CHN-O-RAPID instrument. IR spectra were recorded with a Shimadzu IR-435 spectrometer. The ¹H- and ¹³C-NMR spectra (internal Me4Si) were obtained variously with Varian EM-390 and XL-300, and Brucker AM-200 and WP-80-SY spectrometers. Proximities were established conventionally on the basis of n.O.e. effects. Analytical TLC was preformed on Silica Gel 60 F₂₅₄ (Merck). Silica Gel 60 (230-400 mesh) (Merck) was used for flash-column chromatography.

4-N-Monomethoxytrityl-1-[3',5'-O-(tetraisopropyldisiloxan-1,3-diyl)- β -D-*erythro*-pento-furanos-2'-ulosyl]cytosine (1b)

To a solution of CrO₃ (1.2 g, 12 mmol), pyridine (2 mL, 24 mmol), acetic anhydride (1.2 mL, 12 mmol) and methylene chloride (28 mL), 4-*N*-monomethoxytrityl-1-[3',5'-*O*-(tetraisopropyldisiloxan-1,3-diyl)- β -D-ribofuranosyl]cytosine¹² (3.0 g, 4 mmol) was added. The resulting mixture was stirred at room temperature for 45 min, evaporated to dryness, and the residue, suspended in ethyl acetate (10 mL), was filtered through a silica gel column (20 g) using ethyl acetate as eluent. The filtered solution was evaporated to dryness and the residue coevaporated with ethanol (3 x 5 mL) to give **1b** (2.17 g, 72%) as a white foam. IR (KBr) 1770 cm⁻¹ (C=O furanosulose); ¹HNMR (CDCl₃, 90 MHz): δ 0.8-1.1 (m, 28H, isopropyl), 3.7 (s, 3H, OCH₃), 3.8-4.2 (m, 4H, H-3', H-4', 2H-5'), 4.7 (s, 1H, H-1'), 5.0 (d, 1H, H-5), 5.2 (d. 1H, H-6), 6.7-7.3 (m, 14H, Ar-H). Anal. Calcd. for C4₁H₅₄N₃O₇Si₂: C, 65.04; H, 7.19; N, 5.55. Found: C, 65.33; H, 7.03; N, 5.34.

General procedure for the synthesis of 2'-C-Cyano-2'-deoxy-3',5'-O-(tetraisopyl-disiloxan-1,3-diyl)- β -D-arabinofuranosyl Nucleosides (4)

A mixture of the 2'-ketonucleoside 1 (1 mmol), water (4 mL), ethyl ether (8 mL), sodium bicarbonate (0.16 g, 2 mmol) and sodium cyanide (0.05 g, 1 mmol) was stirred vigorously at room temperature overnight. The organic phase was separated, and the aqueous phase was washed with ethyl ether (2 x 8 mL). The combined ethereal phases were dried over Na₂SO₄, filtered, and evaporated to dryness. The residue, a mixture of two epimeric cyanohydrins (2 and 3), was dissolved in dry acetonitrile (10 mL) to this solution were added 4-(dimethylamino)pyridine (0.25 g, 2 mmol) and (phenyloxy) thiocarbonyl chloride (0.2 mL, 1.1 mmol). The mixture was stirred at room temperature for 2 h, and the solvent was evaporated to dryness. The residue, suspended in toluene (20 mL), was transferred to a three-necked flask. Argon was bubbled through the suspension for 15 min. and then AIBN (0.03 g, 0.2 mmol) and tributyltinhydride (0.4 mL, 1.5 mmol) were added. The flask was heated in an oil bath at 80°C for 1-5 h, while the Argon bubbling was maintained. The reaction mixture was allowed to reach room temperature then, it was washed with water (20 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. The residue was purified by column chromatography.

$1-[2^{\circ}-C-Cyano-2^{\circ}-deoxy-3^{\circ},5^{\circ}-O-(tetraisopropyldisiloxan-1,3-diyl)-\beta-D-arabinofuranosyl]$ uracil (4a)

The residue was chromatographed with ethyl acetate/hexane (1:2) as the eluent to give 0.29 g (40%) of 4a as a white foam. IR (KBr) 2230 cm⁻⁻¹ (CN); ¹HNMR (CDCl₃, 200 MHz): δ 1.02-1.11 (m, 28H, isopropyl), 3.60 (dd, 1H, J_{1',2'}=7.1, J_{2',3'}=9.5 Hz, H-2'), 3.82 (dt, 1H, H-4'), 4.03 (dd, 1H, J_{4',5'a}=2.8, J_{5'a,5'b}=13.4 Hz, H-5a), 4.17 (dd, 1H, J_{4',5'b}=2.4 Hz, H-5'b), 4.65 (dd, 1H, J_{3',4'}=8.4 Hz, H-3'), 5.78 (dd, 1H, J_{5,6}=8.3, J_{5,NH}=1.6 Hz, H-5), 6.29 (d, 1H, H-1'), 7.66 (d, 1H, H-6); ¹³CNMR (CDCl₃, 50 MHz): d 42.15 (C-2'), 59.55 (C-5'), 71.84 (C-3'), 82.04, 84.17 (C-1', C-4'), 103.14 (C-5), 115.51 (CN), 138.28 (C-6), 149.89 (C-2), 162.44 (C-4). Anal. Calcd. for C₂₂H₃₇N₃O₆Si₂: C, 53.30; H, 7.52; N, 8.48. Found: C, 53.15; H, 7.46; N, 8.35.

4-N-Monomethoxytrityl-1-[2'-C-Cyano-2'-deoxy-3',5'-O-(tetraisopropyldisiloxan-1,3-diyl)- β -D-arabinofuranosyl]cytosine (4b)

The residue was chromatographed with ethyl acetate/hexane (1:1) as the eluent to give **4b** (0.32 g, 42%) as a white foam. IR (Nujol) 2225 cm⁻⁻¹ (CN); ¹HNMR (CDCl₃, 300 MHz): δ 0.9-1.1 (m, 28H, isopropyl), 3.66 (t, 1H, J_{1',2'}=J_{2',3'}=7.5 Hz, H-2'), 3.80 (m, 4H, OCH₃, H-4'), 4.03 (m, 2H, 2H-5'), 4.55 (t, 1H, J_{3'},4'=7.5 Hz, H-3'), 5.12 (d, 1H, H-5), 6.29 (d, 1H, H-1'), 6.85 (d, 2H, m-H of MMTr), 7.14-7.37 (m, 12H, Ar-H), 7.40 (d, 1H, H-6); ¹³CNMR (CDCl₃, 50 MHz): δ 42.56 (C-2'), 55.23 (OCH₃), 60.60 (C-5'), 73.37 (C-3'), 82.89, 84.05 (C-1', C-4'), 95.12 (C-5), 115.80 (CN), 139.40 (C-6). Anal. Calcd. for C_{42H57}N₄O₆Si₂: C, 65.76; H, 7.10; N, 7.30. Found: C, 65.84; H, 7.31; N, 7.42.

9-[2'-C-Cyano-2'-deoxy-3',5'-O-(tetraisopropyldisiloxan-1,3-diyl)-β-D-arabinofuranosyl] adenine (4c)

The residue was chromatographed with ethyl acetate/hexane (1:1) to yield 4c (0.23 g, 45%) as a white foam. IR (KBr) 2220 cm⁻⁻¹ (CN); ¹HNMR (CDCl₃, 300 MHz): δ 1.10-1.30 (m, 28H, isopropyl), 3.86 (dd, 1H, J_{1',2'}=7.5, J_{2',3'}=8.5 Hz, H-2'), 4.01 (m, 1H, H-4'), 4.15 (dd, 1H, J_{4',5'a}=2.9, J_{5'a,5'b}=12.7 Hz, H-5'a), 4.32 (dd, 1H, J_{4',5'b}=4.2 Hz, H-5'b), 5.38 (t, 1H, J_{3',4'}=8.5 Hz, H-3'), 6.06 (bs, 2H, NH₂), 6.56 (d, 1H, H-1'), 8.16 (s, 1H, H-2), 8.43 (s, 1H, H-8); ¹³CNMR (CDCl₃, 50 MHz): δ 45.12 (C-2'), 60.99 (C-5'), 73.46 (C-3'), 81.42, 84.43 (C-1', C-4'), 115.29 (CN), 119.93 (C-5), 138.29 (C-8), 149.25 (C-4), 153.15 (C-2), 155.72 (C-6). Anal. Calcd. for C_{23H38}N₆O4Si₂: C, 53.25; H, 7.38; N, 16.20. Found: C, 53.01; H, 7.30; N, 16.41.

Deprotection of Tetraisopropyldisiloxane Group. General Procedure:

To a solution of the silyl-protected nucleosides 4a-c (1 mmol) in THF (15 mL) were added (20 mL, 2 mmol) of a 1M Bu₄NF in THF and the mixture was stirred at room temperature from 15 min. to 3 h. The mixture was evaporated to dryness and the residue was purified by flash column chromatography using ethyl acetate/methanol (10:1) as the eluent to afford the free nucleoside.

1-(2'-C-Cyano-2'-deoxy-β-D-arabinofuranosyl)uracil (5a)

Following the general procedure, **4a** (0.25 g, 0.5 mmol) reacted with Bu₄NF for 15 min, to give, after purification, **5a** (0.04 g, 30%) as a white foam. IR (KBr) 2220 cm⁻¹ (CN); ¹HNMR [(CD₃)₂ SO, 300 MHz]: δ 3.56-3.74 (m, 3H, H-4', 2H-5'), 3.87 (t, 1H, J_{1',2'}=J_{2',3'}=7.8 Hz, H-2'), 4.40 (m, 1H, J_{3',4'}=7.8 Hz, H-3'), 5.20 (t, 1H, OH), 5.69 (d, 1H, J_{5,6}=8.1 Hz, H-5), 6.19 (m, 2H, H-1', OH), 7.93 (d, 1H, H-6). Anal. Calcd. for C₁₀H₁₁N₃O₅: C, 47.43; H, 4.38; N, 16.59. Found: C, 47.28; H, 4.11; N, 16.68.

9-(2'-C-Cyano-2'-deoxy-β-D-arabinofuranosyl)adenine (5c)

According to the general procedure 4c (0.21 g, 0.4 mmol) reacted with Bu₄NF for 1.5 h, to afford, after purification 0.066 g (60%) of **5c** as a white foam. IR (KBr) 2220 cm⁻¹ (CN); ¹HNMR [(CD₃)₂ SO, 50 MHz]: δ 3.45-3.85 (m, 3H, H-4', 2H-5'), 4.07 (dd, 1H, J_{1',2}'=7.1, J_{2',3}'=8.4 Hz, H-2'), 4.86 (m, 1H, J_{3',4}'=8.4 Hz, H-3'), 5.16 (t, 1H, OH), 6.26 (d, 1H, OH), 6.52 (d, 1H, H-1'), 7,34 (bs, 2H, NH₂), 8.16 (s, 1H, H-2), 8.39 (s, 1H, H-8); ¹³CNMR [(CD₃)₂SO, 50 MHz] δ 42.61 (C-2'), 59.64 (C-5'), 71.53 (C-3'), 81.33, 85.29 (C-1', C-4'), 117.14 (CN), 118.76 (C-5), 139.00 (C-8), 148.89 (C-4), 152.86 (C-2), 156.05 (C-6). Anal. Calcd. for C₁₁H₁₂N₆O₃: C, 47.82; H, 4.38; N, 30.42. Found: C, 47.65; H, 4.45; N, 30.53.

2',5'-Bis-O-(tert-butyldimethylsilyl)-4-N-monomethoxytrityl-cytidine (6b)

2',5'-Bis-O-(*tert*-butyldimethylsilyl)cytidine²⁴ (8.00 g, 16.9 mmol) was dissolved in dry CH₂Cl₂ (80 mL) and treated with MMTrCl (10.48 g, 33.9 mmol) and DMAP (83 mg, 0.68 mmol). The mixture was stirred until it had become homogeneous and then Et₃N (4.69 mL, 33.8 mmol) was added. After the mixture being stirred for 1 h, the reaction was quenched by addition of (1:1) pyridine-water (50 mL). Then, the mixture was extracted with CH₂Cl₂ (2 x 50 mL), dried over Na₂SO₄, filtered and evaporated under reduced pressure. The residue was chromatographed with ethylacetate/hexane (1:2) as the eluent to yield 12 g of **6b** (96%) as a yellow foam. ¹HNMR (CDCl₃, 90 MHz): δ 0.8 (s, 18H, *t*-Bu), 2.4 (bs, 1H, OH), 3.6-4.1 (m, 5H, H-2', H-3', H-4', 2H-5'), 3.7 (s, 3H, OCH₃), 4.9 (d, 1H, H-5), 5.9 (d, 1H, J_{1',2}=2 Hz, H-1'), 6.7 (d, 2H, m-H of MMTr), 7.0-7.3 (m, 12H, Ar-H), 7.7 (d, 1H, H-6). Anal. Calcd. for C₄₁H₅₈N₃O₆Si₂: C, 66.09; H, 7.85; N, 5.64. Found: C, 66.12; H, 7.80; N, 5.90.

2',5'-Bis-O-(tert-butyldimethylsilyl)-3'-deoxyuridine (7a)

To a solution of $6a^{16}$ (3.31 g, 7 mmol) in toluene (80 mL), N,N'-thiocarbonyldiimidazol (1.87 g, 10.5 mmol) was added. The mixture was heated at 80°C for 3 h, and then, ethyl acetate (150 mL) and water (75 mL) were added. The organic layer was separated, washed with water (3 x 75 mL), dried over anhydrous sodium sulphate, filtered and evaporated to dryness. The residue, suspended in toluene (100 mL), was transferred to a three-necked flask, AIBN (0.23 g, 1.4 mmol) was added, Argon was bubbled through the suspension for 15 min, and then, tributyltin hydride (7.4 mL, 28 mmol) was added. The flask was heated in an oil bath at 80°C for 5 h, while Argon bubbling was maintained. The reaction mixture was allowed to reach room temperature, and the solvent, evaporated to dryness. The residue was purified by column chromatography with ethyl acetate/hexane (1:3) as the eluent to give 2.33 g (73%) of 7a as white foam. ¹HNMR (CDCl₃, 90 MHz): δ 1.7-2.0 (m, 2H, 2H-3'), 3.7 (dd, 1H, J_{4',5'a}=2, J_{5'a,5'b}=12 Hz, H-5'a), 4.1 (dd, 1H, J_{4',5'b}=2 Hz, H-5'b), 4.1-4.5 (m, 2H, H-2', H-4'), 5.5 (d, 1H, H-5), 5.6 (s, 1H, H-1'), 8.0 (d, 1H, H-6), 9.2 (bs, 1H, NH). Anal. Calcd. for C_{21H40}N₂O₅Si₂: C, 55.20; H, 8.83; N, 6.13. Found: C, 54.93: H, 8.75; N, 6.33.

2',5'-Bis-O-(tert-butyldimethylsilyl)-3'-deoxy-4-N-monomethoxytrityl-cytidine (7b)

The nucleoside **6b** (5.21 g, 7 mmol) dissolved in toluene (80 mL) was treated with N,N'thiocarbonyldiimidazol and AIBN as described for the synthesis of **7a**. The residue obtained, after the work-up, was purified by column chromatography with ethyl acetate/hexane (1:3) to afford 2.65 g (52%) of **7b** as a white foam. ¹HNMR (CDCl₃, 90 MHz): δ 0.8 (s, 18H, t-Bu), 1.5-1.7 (m, 1H, H-3'a), 1.8-1.9 (m, 1H, H-3'b), 3.6 (d, 1H, J_{5'a,5'b}=12 Hz, H-5'a), 4.1 (d, 1H, H-5'b), 4.4 (d, 1H, J_{2',3'}=3 Hz, H-2'), 4.5 (m, 1H, H-4'), 4.9 (d, 1H, H-5), 5.8 (s, 1H, H-1'), 6.8 (d, 2H, m-H of MMTr), 7.1-7.3 (m, 12H, Ar-H), 8.0 (d, 1H, H-6). Anal. Calcd. for C₄₁H₅₈N₃O₅Si₂: C, 67.54; H, 8.02; N, 5.76. Found: C, 67.87; H, 8.12; N, 5.60.

2'-O-(tert-Butyldimethylsilyl)-3'-deoxy-5'-O-monomethoxytrityl-adenine (7d)

To a solution of the protected nucleoside $6d^{16}$ (4.58 g, 7 mmol) in DMF (70 mL), N,N'thiocarbonyldiimidazol (3.74 g, 21 mmol) was added. After stirring at room temperature for 5 h. The reaction was treated with AIBN as described for the synthesis of 7a. The residue obtained, after the work-up, was purified by column chromatography with ethyl acetate/hexane (1:2) as the eluent to give 7d (3.34 g, 75%) as a white foam. ¹HNMR (CDCl₃, 90 MHz): δ 1.8-2.2 (m, 2H, 2H-3'), 3.4-3.5 (m, 2H, 2H-5'), 3.7 (s, 3H, OCH₃), 4.5-4.8 (m, 2H, H-2', H-4'), 5.8 (bs, 2H, NH₂), 5.9 (s, 1H, H-1'), 6.6 (d, 2H, m-H of MMTr), 7.2-7.5 (m, 12H, Ar-H), 8.0 (s, 1H, H-2), 8.3 (s, 1H, H-8). Anal Calcd. for C₃₆H₄₃N₅O₄Si: C, 67.79; H, 6.79; N, 10.98. Found: C, 67.59; H, 6.66; N, 10.75.

General Procedure for Removal of the Protecting Groups of the 2',5'-Bis-O-Silyl-Protected 3'-deoxynucleosides 7

To a solution of the silyl-protected nucleoside 7 (1 mmol) in THF (15 mL) a 1M solution Bu₄NF (20 mL, 2 mmol) was added, and the mixture was stirred at room temperature for 1-3 h. The reaction mixture was filtered to through a wet (THF) column of silica gel using THF as the eluent. The filtrate was evaporated to dryness, and the residue was purified by column chromatography with ethyl acetate/methanol (10:1) as the eluent to afford the free nucleoside.

3'-Deoxyuridine (8a)¹⁷

Following the general procedure 7a (1.37 g, 3 mmol) reacted with Bu₄NF for 3 h, to yield, after purification, 0.66 g (96%) of 8a as a white foam. ¹HNMR [(CD₃)₂SO, 90 MHz]: δ 1.6-1.9 (m, 2H, 2H-3'), 3.4-3.6 (m, 2H, 2H-5'), 3.9-4.3 (m, 2H, H-2', H-4'), 4.9 (t, 1H, OH), 5.3-5.6 (m, 3H, H-1', H-5, OH), 7.8 (d, 1H, H-6). Anal. Calcd. for C₉H₁2N₂O₅: C, 47.36; H, 5.30; N, 12.28. Found: C, 47.68; H, 5.39; N, 12.03.

3'-Deoxy-4-N-monomethoxytrityl-cytidine (8b)

According to the general procedure **7b** (2.62 g, 3.6 mmol) reacted with Bu₄NF for 2 h, to give 1.25 (70%) of **8b** as a white foam. ¹HNMR [(CD₃)₂SO, 90 MHz]: δ 1.6-1.9 (m, 2H, 2H-3'), 3.5-3.7 (m, 2H, 2H-5'), 3.7 (s, 3H, OCH₃), 3.9-4.3 (m, 3H, H-2', H-4', H-5), 5.0 (t, 1H, OH), 5.4 (d, 1H, OH), 5.6 (d, 1H, J_{1',2'}=2 Hz, H-1'), 6.9 (d, 2H, m-H of MMTr), 7.1-7.4 (m, 12H, Ar-H), 7.9 (d, 1H, H-6), 8.3 (bs, 1H, NH). Anal. Calcd. for C₂₉H₂₉N₃O₅: C, 69.72; H, 5.85; N, 8.41. Found: C, 69.98; H, 5.96; N, 8.60.

3'-Deoxy-5'-O-monomethoxytrityl-adenosine (8d)¹⁸

Following the general procedure 7d (3.19 g, 5 mmol) reacted with 1 eq of Bu₄NF (50 mL, 5 mmol) for 1 h. To give, after purification, 2.4 g (93%) of 8d as a white foam. ¹HNMR (CDCl₃, 90 MHz): δ 2.0-2.2 (m, 2H, 2H-3'), 3.2-3.4 (m, 2H, 2H-5'), 3.7 (s, 3H, OCH₃), 4.6-4.8 (m, 2H, H-2', H-4'), 5.9 (d, 1H, J₁',₂'=2 Hz, H-1'), 6.3 (bs, 2H, NH₂), 6.7 (d, 2H, m-H of MMTr), 7.1-7.4 (m, 12H, Ar-H), 8.0 (s, 1H, H-2), 8.2 (s, 1H, H-8). Anal. Calcd. for C₃₀H₂₉N₅O₄: C, 68.22; H, 5.58; N, 13.38. Found: C, 68.40; H, 5.64; N, 13.23.

5'-O-(tert-Butyldimethylsilyl)-3'-deoxyuridine (9a)

To an ice cooled solution of **8a** (0.66 g, 2.9 mmol) in dry pyridine (30 mL) *tert*-butyldimethylsilyl chloride (0.44 g, 2.9 mmol) was added, in portions, during 2 h. The resulting mixture was stirred at room temperature overnight, and then, evaporated to dryness. The residue, dissolved in chloroform, was washed with water (2 x 15 ml) dried over Na₂SO₄, filtered and evaporated to dryness. The residue was purified by column chromatography with ethyl acetate/methanol (30:1) as the eluent to yield 0.71 g (72%) of **9a** as a white foam. ¹HNMR (CDCl₃, 90 MHz): δ 0.9 (s, 9H, t-Bu), 1.8-2.1 (m, 2H, 2H-3'), 3.7 (dd, 1H, J_{4',5'a=1}, J_{5'a,5'b=12}

Hz, H-5'a), 4.2 (dd, 1H, $J_{4',5'b}=1$ Hz, H-5'b), 4.4-4.7 (m, 3H, H-2', H-4', O-H), 5.6 (d, 1H, H-5), 5.7 (s, 1H, H-1'), 8.2 (d, 1H, H-6). Anal. Calcd. for $C_{15}H_{26}N_2O_5Si: C, 52.60; H, 7.65; N, 8.18$. Found: C, 52.30; H, 7.48; N, 8.03.

5'-O-(tert-Butyldimethylsilyl)-3'-deoxy-4-N-monomethoxytrityl-cytidine (9b)

Compound **8b** (1.25 g, 2.5 mmol) reacted with *tert*-butyldimethylsilylchloride (0.38 g, 2.5 mmol) as described for the synthesis of **9a**. After the work-up, the residue was purified by column chromatography with ethyl acetate/methanol (20:1) to give 1.24 g (81%) of **9b** as a white foam. ¹HNMR [(CD₃)₂SO, 90 MHz]: δ 0.9 (s, 9H, t-Bu), 1.7-1.9 (m, 2H, 2H-3'), 3.7 (s, 3H, OCH₃), 3.6-4.4 (m, 5H, H-2', H-4' 2H-5', H-5), 5.5 (bs, 1H, OH), 5.6 (s, 1H, H-1'), 6.9 (d, 2H, m-H of MMTr), 7.1-7.4 (m, 12H, Ar-H), 7.8 (d, 1H, H-6), 8.4 (bs, 1H, NH). Anal. Calcd. for C₃₅H₄₄N₃O₅Si: C, 68.37; H, 7.21; N, 6.83. Found: C, 68.00; H, 7.03; N, 6.97.

1-[5'-O-(tert-butyldimethylsilyl)-3'-deoxy-β-D-glycero-pentofuranos-2'-ulosyl]uracil (10a)

Compound **9a** (0.68 g, 2 mmol) was added to a solution of CrO₃/pyridine/acetic anhydride as described for the synthesis of **1b**. After the work-up 0.44 g (65%) of **10a** was obtained, as a yellow syrup. IR (Nujol) 1770 cm⁻¹ (C=O furanosulose); ¹HNMR (CDCl₃, 200 MHz): δ 0.90 (s, 9H, t-Bu), 2.65 (dd, 1H, J_{3'a,4'=7.5}, J_{3'a,3'b=18.6 Hz, H-3'a), 2.85 (dd, 1H, J_{3'b,4'=7.6 Hz, H-3'b), 3.86 (dd, 1H, J_{4',5'a=4.4}, J_{5'a,5'b=11.1 Hz, H-5'a), 3.93 (dd, 1H, J_{4',5'b=3.9} Hz, H-5'b), 4.51 (m, 1H, H-4'), 5.50 (s, 1H, H-1'), 5.72 (d, 1H, H-5), 7.26 (d, 1H, H-6), 9.47 (bs, 1H, NH). Anal. Calcd. for C₁₅H₂₄N₂O₅Si: C, 52.91; H, 7.10; N, 8.23. Found: C, 52.59; H, 7.31; N, 8.03.}}}

4-N-Monomethoxytrityl-1-[5'-O-(*tert*-butyldimethylsilyl)-3'-deoxy- β -D-glycero-pento-furanos-2'-ulosyl]cytosine (10b)

Following the oxidation procedure described for the synthesis of 1b, compound 9b (1.23 g, 2 mmol) reacted with CrO₃/pyridine/acetic anhydride for 1 h, to afford, after the work-up 1.08 g (89%) of 10b as a whitte foam. IR (Nujol) 1775 cm⁻¹ (C=O furanosulose); ¹HNMR (CDCl₃, 90 MHz): δ 0.9 (s, 9H, t-Bu), 2.4 (dd, 1H, J_{3'a,4'}=6.0, J_{3'a,3'b}=18.0 Hz, H-3'a), 2.9 (dd, 1H, J_{3'b,4'}=8.0 Hz, H-3'b), 3.7 (s, 3H, OCH₃), 3.8 (d, 2H, 2H-5'), 4.4 (m, 1H, H-4'), 5.0 (m, 2H, H-1', H-5), 6.7-7.3 (m, 15H, ArH, H-6). Anal. Calcd for C₃₅H₄₂N₃O₅Si: C, 68.59; H, 6.91; N, 6.86. Found: C, 68.29; H, 6.71; N, 6.79.

9-(3'-Deoxy-5'-O-monomethoxytrityl-β-D-glycero-pentofuranos-2'-ulosyl)adenine (10d)

According to the procedure described for the synthesis of **1b**, compound **8d** (1.05 g, 2 mmol) reacted with CrO3/pyridine/acetic anhydride for 45 min, to give 0.55 g (53%) of **10d** as a brown foam. IR (Nujol) 1770 cm⁻¹ (C=O furanosulose); ¹HNMR (CDCl₃, 90 MHz): d 2.7 (dd, 1H, $J_{3'a,4'}=7.0$, $J_{3'a,3'b}=18.0$ Hz, H-3'a), 3.2 (dd, 1H, $J_{3'b,4'}=7.0$ Hz, H-3'b), 3.4 (d, 2H, 2H-5'), 3.7 (s, 3H, OCH₃), 4.5-4.7 (m, 1H, H-4') 5.7 (s, 1H, H-1'), 6.0 (bs, 2H, NH₂), 6.7 (d, 2H, m-H of MMTr) 7.1-7.4 (m, 12H, Ar-H), 7.8 (s, 1H, H-2), 8.2 (s, 1H, H-8). Anal. Calcd. for $C_{30}H_{27}N_5O_4$: C, 69.08; H, 5.22; N, 13.43. Found: C, 69.40; H, 5.13; N, 13.59.

1-[5'-O-(*tert*-Butyldimethylsilyl)-2'-C-cyano-2',3'-dideoxy-β-D-*threo*-pentofuranosyl] uracil (13a)

According to the general procedure described for the synthesis of 2'-C-cyano-2'-deoxynucleosides (4), the 2'-ketonucleoside **10a** (0.41 g, 1.2 mmol), reacted with NaCN overnight, then with (phenyloxy)thiocarbonyl chloride and finally with Bu₃SnH for 3 h, to give, after the work-up a residue which was chromatographed with ethyl acetate/hexane (1:2) to afford 0.21 g (50%) of **13a** as a white foam. IR (KBr) 2225 cm⁻¹ (CN); ¹HNMR (CDCl₃, 200 MHz): δ 0.87 (s, 9H, t-Bu), 2.30 (m, 2H, 2H-3'), 3.59 (m, 1H, H-2'), 3.69 (dd, 1H, J_{4',5'a}=2.6,

 $J_{5'a,5'b}=11.7$ Hz, H-5'a), 3.99 (dd, 1H, $J_{4',5'b}=2.8$ Hz, H-5'b), 4.1 (m, 1H, H-4'), 5.68 (d, 1H, H-5), 6.20 (d, 1H, $J_{1',2'}=8.7$ Hz, H-1'), 7.92 (d, 1H, H-6); ¹³CNMR (CDCl₃, 50 MHz): δ 25.94 (CH₃), 29.45 (C-3'), 35.30 (C-2'), 62.19 (C-5'), 84.06, 81.04 (C-1', C-4'), 102.76 (C-5), 116.47 (CN), 139.18 (C-6), 150.31 (C-2), 162.89 (C-4). Anal. Calcd. for C₁₆H₂₅N₃O₄Si: C, 54.67; H, 7.17; N, 11.95. Found: C, 54.39; H, 7.06; N, 12.05.

1-[5'-O-(*tert*-butyldimethylsilyl)-2'-C-cyano-2',3'-dideoxy-b-D-*threo*-pentofuranosyl]-4-Nmonomethoxytrityl-cytosine (13b)

The 2'-ketonucleoside **10b** (0.67 g, 1,1 mmol) reacted with NaCN/(phenyloxy) thiocarbonyl chloride/Bu₃SnH, following the general procedure described for the synthesis of 2'-C-cyano-2'-deoxynucleosides (4). After the work-up, the residue was purified by column chromatography with ethylacetate, as the eluent, to give **13b** (0.24 g, 36%) as a white foam. IR (KBr) 2220 cm⁻⁻¹ (CN); ¹HNMR (CDCl₃, 300 MHz): δ 0.78 (s, 9H, t-Bu), 2.27-2.34 (m, 2H, 2H-3'), 3.65-3.73 (m, 2H, H-2', H-5'a), 3.78 (s, 3H, OCH₃), 3.80 (dd, 1H, J_{4',5'b}=3.6, J_{5'a,5'b}=11.4 Hz, H-5'b), 4.13 (m, 1H, H-4'), 5.05 (d, 1H, H-5), 6.19 (d, 1H, J_{1',2'}=6.2 Hz, H-1'), 6.80-6.84 (d, 2H, m-H of MMTr), 7.10-7.32 (m, 12H, Ar-H), 7.63 (d, 1H, H-6); ¹³CNMR (CDCl₃, 50 MHz): δ 29.94 (C-3'), 34.76 (C-2'), 55.20 (OCH₃) 62.77 (C-5'), 79.85-84.87 (C-1', C-4'), 116.91 (CN). Anal. Calcd. for C₃₆H₄₂N₄O₄Si: C, 69.42; H, 6.80; N, 9.00. Found: C, 69,10; H, 6.70; N, 8.88.

9-(2'-C-Cyano-2',3'-dideoxy-5'-O-monomethoxytrityl-β-D-threo-pentofuranosyl)adenine (13d)

The 2'-ketonucleoside **10d** (0.52 g, 1.0 mmol) reacted with NaCN/(phenyloxy) thiocarbonyl chloride/Bu₃SnH, according to the general procedure described for the synthesis of **4**. The residue was purified by column chromatography with ethyl acetate/hexane (2:1) to give **13d** (0.13 g, 25%) as a white foam. IR (KBr) 2220 cm⁻⁻¹ (CN); ¹HNMR (CDCl₃, 300 MHz): δ 2.51 (m, 1H, H-3'a), 2.64 (m, 1H, H-3'b), 3.48 (d, 2H, 2H-5'), 3.70 (m, 1H, H-2'), 3.78 (s, 3H, OCH₃), 4.30 (m, 1H, H-4'), 5.89 (bs, 2H, NH₂), 6.48 (d, 1H, J_{1',2'}=6.7 Hz, H-1'), 6.84 (d, 2H, m-H of MMTr), 7.2-7.4 (m, 12H, Ar-H), 8.17 (s, 1H, H-2), 8.30 (s, 1H, H-8); ¹³CNMR (CDCl₃, 50 MHz): δ 31.39 (C-3'), 35.45 (C-2'), 55.23 (OCH₃) 63.89 (C-5'), 79.57-83.60 (C-1', C-4'), 116.25 (CN). Anal. Calcd. for C₃₁H₂₈N₆O₃: C, 69.91; H, 5.30; N, 15.78. Found: C, 70.24; H, 5.21; N, 15.97.

1-(2'-C-Cyano-2',3'-dideoxy-β-D-threo-pentofuranosyl)uracil (14a)

The protected nucleoside 13a (0.2 g, 0.6 mmol) was stirred with methanolic 0.1 M HCl (4 mL) at room temperature for 45 min. The solution was neutralized with 1M NaOH-MeOH and the solvent was evaporated to dryness. The residue was purified by column chromatography using dichloromethane/methanol (20:1) as the eluent to afford 0.12 g (86%) of 14a as white foam. IR (KBr) 2220 cm⁻¹ (CN); ¹HNMR [(CD₃)₂SO, 200 MHz]: δ 2.13 (dd, 1H, H-3'a), 2.39 (m, 1H, H-3'b), 3.56 (m, 1H, J_{4',5'a}=J_{5'a,5'b}=10 Hz, H-5'a), 3.75 (m, 1H, J_{4',5'b}=2 Hz, H-5'b), 4.02 (m, 2H, H-2', H-4'), 5.20 (bs, 1H, OH), 5.67 (d, 1H, H-5), 6.15 (d, 1H, J_{1',2'}=7.1 Hz, H-1'), 8.03 (d, 1H, H-6), 10.6 (bs, 1H, NH). ¹³CNMR [(CD₃)₂SO, 50 MHz): δ 29.25 (C-3'), 34.21 (C-2'), 60.26 (C-5'), 81.42-83.51 (C-1', C-4'), 101.62 (C-5), 117.81 (CN), 139.80 (C-6), 150.19 (C-2), 162.94 (C-4). Anal. Calcd. for C₁₀H₁₁N₃O₄: C, 50.63; H, 4.67; N, 17.71. Found: C, 50.55; H, 4.63; N, 17.60.

1-(2'-C-Cyano-2',3'-dideoxy-β-D-threo-pentofuranosyl)cytosine (14c)

The protected nucleoside 13b (0.37 g, 0.6 mmol) was stirred with methanolic 0.1M HCl (8 mL) at room temperature for 48 h. The reaction was neutralized with 1M NaOH-MeOH and the solvent was evaporated to

dryness. The residue was purified by column chromatography with dichloromethane/methanol (5:1), as the eluent, to yield 0.11 g (80%) of 14c as white foam. IR (KBr) 2220 cm⁻¹ (CN); ¹HNMR [(CD₃)₂SO 300 MHz]: δ 2.03-2.14 (m, 1H, H-3'a), 2.32-2.41 (m, 1H, H-3'b), 3.34 (s, 3H, OCH₃), 3.50-3.60 (m, 1H, J_{5'a,5'b}=12.3 Hz, H-5'a), 3.65-3.76 (m, 1H, J_{4',5'b}=3.2 Hz, H-5'b), 3.93-4.02 (q, 1H, J_{1',2'}=J_{2',3'a}=J_{2',3'b}=7.1 Hz, H-2'), 4.03-4.11 (m, 1H, H-4'), 5.15 (t, 1H, OH), 5.78 (d, 1H, H-5), 6.12 (d, 1H, H-1'), 7.20 (d, 2H, NH₂), 7.92 (d, 1H, H-6). ¹³CNMR [(CD₃)₂SO, 50 MHz): δ 29.84 (C-3'), 34.23 (C-2'), 60.72 (C-5'), 80.68 (C-4'), 84.10 (C-1'), 94.10 (C-5), 118.27 (CN), 140.53 (C-6), 155.07 (C-2), 165.66 (C-4). Anal. Calcd. for C₁₀H₁₂N₄O₃: C, 50.84; H, 5.12; N, 23.72. Found: C, 50.70; H, 4.95; N, 27.98.

9-(2'-C-Cyano-2',3'-dideoxy-β-D-threo-pentofuranosyl)adenine (14d)

The protected nucleoside 13d (0.13 g, 0.2 mmol) was dissolved in 80% acetic acid solution (2.4 mL) and the reaction mixture was heated at 90°C for 30 min. The solvent was evaporated to dryness. The residue was coevaporated with ethanol (2 x 2 mL) and purified by column chromatography with dichloromethane/methanol (10:1) to afford 14d (0.40 g, 70%) as a white foam. IR (KBr) 2220 cm⁻⁻¹ (CN); ¹HNMR [(CD₃)₂SO, 300 MHz]: δ 2.45-2.55 (m, 2H, 2H-3'), 3.60-3.80 (m, 2H, 2H-5'), 4.25-4.30 (m, 2H, H-2', H-4'), 5.25 (t, 1H, OH), 6.52 (d, 1H, J_{1',2'}=7.0 Hz, H-1'), 7.25 (bs, 2H, NH₂), 8.19 (s, 1H, H-2), 8.50 (s, 1H, H-8). Anal. Calcd. for C₁₁H₁₂N₆O₂: C, 50.76; H, 4.65; N, 32.29. Found: C, 50.64; H, 4.59; N, 32.00.

Acknowledgement.—We thank Rhône-Poulenc Farma S.A.E. for financial support. We also thank the Plan Nacional de I + D Farmacéuticos of Spain for a grant to (S. V.).

REFERENCES

- 1. a) Broder, S. Med. Res. Rev. 1990, 10, 419-439; b) Mitsuya, H. Nature, 1987, 325, 773-778.
- 2. Mitsuya, H.; Broder, S. Proc. Natl. Acad. Sci. USA, 1986, 83, 1911-1915.
- 3. Herdewijn, P.; De Clercq, E. Dideoxynucleoside analogues as inhibitors of HIV replication. In Design of Anti-AIDS Drugs; De Clercq, E., Ed.; Elsevier, Amsterdam, 1990; pp. 141-174.
- 4. Fischl, M.A. Richmann, D.D.; Grieco, M.H. et. al. N. Engl. J. Med. 1987, 317, 185-191.
- 5. Richman, D.D.; Fisch, M.A.; Grieco, M.H. et al. N. Engl. J. Med. 1987, 317, 192-197.
- 6. Yaorchan, R.; Klecker, R.W.; Weinhold, K.J. et al. Lancett, 1986, i, 575-580.
- De las Heras, F.G.; Camarasa, M.J.; Fiandor, J. Nucleosides: Potential drugs for AIDS therapy. In Recent Progress in the Chemical Synthesis of Antibiotics; Lukacs, G.; Ohno, M., Eds.; Springer Verlag; Berlin-Heidelberg, 1990, pp. 321-363.
- De Clercq, E.; New developments in Anti-AIDS drug research. In Trends in Drug Research; Chaasen, V.; Ed.; Elsevier Science Publishers; Amsterdam, 1990, pp. 133-152.
- Camarasa, M.J.; Díaz-Ortiz, A.; Calvo-Mateo, A.; De las Heras, F.G. J. Med. Chem. 1989, 32, 1732-1738.
- 10. Calvo-Mateo, A.; Camarasa, M.J.; Díaz-Ortiz, A.; De las Heras, F.G. Tetrahedron Lett. 1988, 29, 941-944.
- 11. Calvo-Mateo, A.; Camarasa, M.J.; Díaz-Ortiz, A.; De las Heras, F.G. Tetrahedron 1988, 44, 4895-4903.
- 12. Honda, S.; Urakami, K.; Terada, K.; Sato, Y.; Kohno, K.; Sekine, M.; Hata, T. Tetrahedron 1984, 40, 153-163.
- 13. Hansske, F.; Madej, D.; Robins, M.J. Tetrahedron, 1984, 40, 125-135.

- 14. Hartwig, W. Tetrahedron, 1983, 39, 2609-2645.
- 15. Robins, M.J.; Wilson, J.S. J. Am. Chem. Soc. 1981, 103, 932-933.
- 16. Ogilvie, K.K.; Beaucage, S.L.; Schifman, A.L.; Theriault, N.Y.; Sadana, K.L. Can. J. Chem. 1978, 56, 2768-2780.
- 17. Novak, J.J.K.; Sorm, F. Collect. Czech. Chem. Commun. 1973, 38, 1173-1178.
- 18. Robins, M.J.; Jones, R.A.; Malcon, McC. Biochemistry 1974, 13, 553-559.
- 19. Bernstein, M.A.; Morton, H.E.; Guidon, Y. J. Chem. Soc., Perkin 2 1986, 1155-1163.
- 20. Rosemeyer, H.; Seela, F. Nucleosides. Nucleotides 1990, 9, 417-418.
- 21. a) Matusda, A.; Takenuki, K.; Itoh, H.; Sasaki, T.; Ueda, T. Chem. Pharm. Bull. 1987, 35, 3967-3970.
 b) Matsuda, A.; Takenuki, K.; Sasaki, T.; Ueda, T. J. Med. Chem. 1991, 34, 234-239.
- a) Robins, M.J.; Mac Coss, M.; Wilson, J.S. J. Am. Chem. Soc. 1977, 99, 4660-4666. b) Robins, M.J.; Wilson, J.S.; Hansske, F. J. Am. Chem. Soc. 1983, 105, 4059-4065.
- 23. Matsuda, A.; Nakajima, Y.; Azuma, A.; Tanaka, M.; Sasaki, T. J. Med. Chem. 1991, 34, 2917-2919.
- 24. Ogilvie, K.K.; Shifman, A.L., Penney, C.L. Can. J. Chem. 1978, 56, 2768-2780.